

1 **TITLE**

2 Circulating anti-human leukocyte antigen IgM antibodies as a potential early predictor of
3 allograft rejection and negative clinical outcome after lung transplantation

4

5 **Authors:**

6 Kazuaki Miyahara, MD¹, Kentaroh Miyoshi, MD, PhD¹, Takeshi Kurosaki MD, PhD², Shinji
7 Otani MD, PhD¹, Seiichiro Sugimoto MD, PhD^{1,2}, Masaomi Yamane MD, PhD¹, Shinichi
8 Toyooka MD, PhD^{1,2}

9

10 **Institution and affiliations:**

11 ¹Department of Thoracic Surgery, Okayama University Hospital, 2-5-1, Shikata-cho, kita-ku,
12 Okayama 700-8558, Japan.

13 ²Organ Transplant Center, Okayama University Hospital, 2-5-1, Shikata-cho, kita-ku, Okayama
14 700-8558, Japan.

15

16 **Article type:**

17 Original article (Clinical Original)

18

19 **Corresponding Author:**

20 Kentaroh Miyoshi MD, PhD,

21 Department of Thoracic Surgery, Okayama University Graduate School of Medicine, Dentistry

22 and Pharmaceutical Science, 2-5-1, Shikata-cho, kita-ku, Okayama 700-8558, Japan.

23 Tel: +81-86-235-7265

24 Fax: +81-86-235-7269

25 E-mail address: kmiyoshi@okayama-u.ac.jp

26

27 **Keywords:**

28 Immunoglobulin M, antibody, rejection, biomarker, lung transplantation

29 **ABSTRACT**

30 **Purposes:** Anti-human leukocyte antigen (HLA) immunoglobulin (Ig) M production stimulated
31 by an alloantigen is sensitive, thus making IgM a potential and novel marker of allorejection
32 after organ transplantation. This study aimed to examine the relationship between serum levels
33 of anti-HLA IgM early after clinical lung transplantation (LTx) and post-transplant outcomes.

34 **Methods:** Thirty-one consecutive patients who underwent deceased LTx were included.
35 Immunoreactivity against HLA was retrospectively analyzed by measuring anti-HLA IgM of the
36 serum sampled for the first 14 days after LTx. The flow panel reactive antibody technique was
37 used. The ratio of the anti-class I IgM level at each day to baseline was obtained, and a peak
38 IgM was determined for each case. Correlation between the peak IgM level and subsequent
39 development of acute rejection (AR), chronic lung allograft dysfunction (CLAD), and survival
40 outcomes were examined.

41 **Results:** The peak IgM level was a significant risk factor for AR within 90 days in univariate
42 and multivariate analyses. In the long term, the patients with positive IgM (peak level > 1.8)
43 tended to have a poorer CLAD-free and overall survival.

44 **Conclusion:** Elevation of anti-HLA IgM levels early after LTx potentially correlated with a
45 higher incidence of rejection and worse clinical outcomes.

46 **MAIN BODY**

47 **Introduction**

48 Despite the recent establishment of a basic methodology for immunosuppressant
49 management, chronic lung allograft dysfunction (CLAD) remains a major limitation to
50 long-term survival for lung transplant (LTx) recipients. According to the international database,
51 bronchiolitis obliterans syndrome (BOS), a typical phenotype of CLAD, is the leading
52 morbidity and cause of mortality for >1-year survivors after LTx [1]. It is recognized that
53 multifactorial events in the early post-transplant period, which are associated with both natural
54 and acquired immunity, are involved in the subsequent chronic rejection process [2-5]. Of these,
55 the alloimmune response provoked by mismatched HLA antigens plays a pivotal role in the
56 development of chronic allograft dysfunction. However, there is no early prognostic predictor or
57 a monitoring method to optimize the personalized adjustment of immunosuppression [6].

58 A growing number of recent studies have focused on donor-specific alloantibody
59 (DSA) as a cause of reluctant antibody-mediated rejection (AMR) and irreversible allograft
60 dysfunction. Most studies have identified the actual impact of DSA and subclinical or clinical
61 AMR on CLAD or patients' survival [7-10]. Post-transplant de novo DSA discussed here was
62 categorized as class immunoglobulin G (IgG) and generally regarded as one of the clinical
63 markers for poor prognosis. However, the timing of problematic elevation of serum de novo
64 DSA-IgG levels reportedly varies widely and is mostly at more than one month after LTx. Once

65 DSA-IgG emerges, furthermore, the elimination and treatment of rejection require painstaking
66 processes and often fail. These facts suggest that DSA-IgG does not necessarily work
67 effectively as an early clinical marker to detect reluctant acute rejection or CLAD in the
68 preventable or treatable stage.

69 We previously reported that donor-specific immunoglobulin M (IgM) levels in the
70 recipient's serum were promptly elevated post-transplant in response to acute rejection in the
71 clinical setting of bilateral living-donor LTx [11]. Production of IgM from plasma cells is
72 triggered by a primitive antigen exposure process. The responses of DSA-IgM were specifically
73 observed in a very early stage, prior to the development of clinical signs of acute rejection (AR)
74 in the study. IgM production is sensitive to immunoreaction and is stimulated theoretically in
75 accordance with the extent of reactivity against donor antigens [11-13]. This hypothesis
76 supports the hypothesis that elevated levels of recipient serum IgM directed to human leukocyte
77 antigen (HLA) can be a sensitive marker of immunoreaction against allograft and early
78 predictor of long-term graft survival after LTx. This study aimed to examine the relationship
79 between serum levels of anti-HLA IgM early after LTx and subsequent post-transplant
80 outcomes.

81

82 **Materials and Methods**

83 **Patients and study design**

84 A total of 31 patients who underwent deceased-donor LTx between July 2013 and July
85 2016 at Okayama University Hospital were included in the study. Plasma samples from these
86 patients were obtained and preserved once pretransplant and on days 2, 4, 7, 10, and 14 after
87 transplantation. In addition, the medical records of the patients were reviewed to ascertain the
88 independent variables determining the clinical characteristics and post-transplant outcomes.
89 Serum samples and clinical data were obtained following approval of the institutional review
90 board of Okayama University Hospital (approval #: 1609-026) and consent from the patients.

91 Statistical analyses were conducted to explore the relationship between patient
92 variables, including acute IgM levels and subsequent acute rejection events within 90 days after
93 LTx. Furthermore, the study cohort was divided into IgM-positive group and IgM-negative
94 groups based on the cutoff level determined by a receiver operating characteristic curve analysis.
95 The incidence of CLAD and the long-term survival were then evaluated and compared between
96 the groups.

97

98

99 **Recipient and Donor selection**

100 Patients who had officially approved indications for LTx were registered on the waitlist
101 provided by the Japan Organ Transplant Network (JOTN). Procedure indication was determined

102 for each candidate according to the primary disease, urgency, and organ availability. The
103 patients in the cohort had no previous sensitized history. Offered deceased-donor lungs were
104 allocated to recipients according to waitlist order, ABO compatibility, matching of predicted
105 pulmonary function value, and negative result of lymphocyte crossmatching test
106 (complement-dependent cytotoxicity test for warm T and B cells). Detailed donor data,
107 including past medical history and examination results, were obtained by authorized donor
108 coordinators and shared among the donor hospital staff and transplant centers with the use of a
109 form filled out by the authorized donor coordinators and transplant physicians [14]. Our
110 institutional expert team made a final decision to accept or decline the donor organs.

111

112 **Transplant procedure and post-transplant management**

113 The lung procurement procedure was standardized for all deceased donors. The
114 donors' lungs were routinely flushed with EP-TU extracellular solution (Cell Science &
115 Technology Institute, Sendai, Japan). LTx surgery was performed in a standard manner via a
116 transsternal anterior thoracotomy approach. Intraoperative cardiopulmonary support with a
117 standard bypass technique was used when the unilateral native lung was not able to maintain
118 adequate gas exchange and hemodynamic stability. Recipients received a standard triple-drug
119 immunosuppressive regimen post-transplant consisting of a calcineurin inhibitor (tacrolimus), a

120 cell-cycle inhibitor (mycophenolate mofetil, MMF), and steroids (prednisolone). The target
121 trough level of tacrolimus was set at 8-11 ng/mL. The doses of MMF and prednisolone were
122 optimized according to the patient's body weight. The prednisolone tapering policy was
123 standardized.

124

125 **Detection of Anti-HLA IgM antibodies after LTx**

126 Recipient serum IgM with an affinity for the HLA panel was retrospectively
127 analyzed at the Central Research Laboratory, Okayama University Medical School. Blood
128 samples were obtained on days 2, 4, 7, 10, and 14 after LTx, and preserved following
129 centrifugal separation. Flow cytometry crossmatching was performed with the use of flow panel
130 reactive antibody (PRA) screening test HLA class I beads (One Lambda) referring to the
131 manufacture's instruction for IgG detection. Briefly, 5 μ L of the beads were mixed with 20 μ L
132 of sera and incubated for 30 min. Then, secondary antibody to IgM (Jackson ImmunoResearch),
133 instead of IgG, was added and reacted to the composites for 30 min to label the anti-class I HLA
134 IgM caught by the panel beads. Fluorescence intensity was analyzed using a MACS Quant
135 Analyzer (Milteny Biotec). The mean fluorescence intensity (MFI) for overall panel beads
136 reacted to the IgM was calculated. All the process required two hours per single examination.

137

138 **Definition for data interpretation**

139 The MFI value of day 2 sample for the flow PRA IgM testing was defined as the
140 baseline anti-HLA IgM level. Then, the ratio of the MFI values on the following post-transplant
141 days to baseline was calculated. Through the paired sera examination, the maximal value of the
142 ratio of the first 14 days after LTx was adopted as representative data for the acute anti-HLA
143 IgM level for each case. AR was diagnosed cautiously by experienced transplant clinicians
144 based on the following clinical findings: (1) elevation of an inflammatory marker (C-reactive
145 protein); (2) deteriorated oxygenation; (3) ground-glass opacities, peribronchovascular
146 infiltration and interlobular septal thickening detected by high-resolution computed tomography;
147 (4) increased serous pleural effusion; and (5) the clinical manifestations above were well
148 responsive to pulse steroid treatments. To determine the pre-transplant severity of each patient,
149 the US lung allocation score (LAS) was retrospectively calculated (in November 2016) using
150 the LAS calculator available on the OPTN website
151 (<https://optn.transplant.hrsa.gov/resources/allocation-calculators/las-calculator/>). Primary graft
152 dysfunction (PGD) grade and the diagnosis of CLAD (BOS and restrictive phenotype) were
153 defined in accordance with the International Society for Heart and Lung Transplantation
154 working group statement [15,16].

155

156 **Statistical Analysis**

157 Cox regression analysis was used to examine the influence of various patient factors
158 on the development of AR within 90 days after LTx and CLAD. The rate of freedom from
159 CLAD and overall survival were depicted using Kaplan–Meier analysis and compared between
160 the IgM-positive and negative groups using the log-rank test. A p-value <0.05 was considered
161 statistically significant. All analyses were performed using SPSS 24.0.

162

163 **Results**

164 **Patient characteristics and anti-HLA antibody levels early after transplantation**

165 Overall, 155 samples from 31 recipients were analyzed to examine anti-HLA IgM
166 levels in the acute phase. The patient characteristics are shown on Table 1. The cohort
167 comprised 15 women and 16 men. The mean age was 38.4 (range, 8–61) years. The indications
168 for transplantation were interstitial lung disease (n = 10, 32.2%), bronchiolitis obliterans (n = 7,
169 22.6%), pulmonary hypertension (n = 3, 9.7%), chronic obstructive pulmonary disease (n = 1,
170 3.2%), and others (n = 6, 19.4%). Ten patients (32.2%) had sensitization history such as
171 pregnancy and blood transfusion. The mean HLA mismatch number was 4.3. Pretransplant
172 MFI levels of anti-HLA IgM ranged from 0.75 to 14.74 (mean, 2.62). In order to contrast the
173 patient's IgM reactivity, we defined the MFI rise rate as the IgM level in each case (MFI value

174 divided by individual baselines). Eventually, peak IgM levels (maximal rise rate) ranged from
175 0.81 to 2.52 (mean, 1.47) in this patient cohort. Transitional changes in the average IgM levels
176 are shown in Figure 1A. There was a general upward trend in the IgM rise rate peaking around
177 10 days after LTx.

178 An example course of anti-HLA IgM transition in a representative case is depicted in
179 the Case1, Figure 2. The patient was a 33-year-old man with obstructive pulmonary disease. He
180 underwent right single LTx and subsequently developed steroid-responsive acute rejection on
181 postoperative day 8. The peak IgM level in this case was regarded as 2.46.

182

183 **Impact of peak anti-HLA IgM level on the development of AR and CLAD**

184 Seven patients developed AR during the first 90 days after LTx. All the IgM data for
185 the patients with AR are demonstrated in Figure 2. The onset of AR was found from sixth to
186 sixtieth posttransplant day (mean 17.43 days after LTx). The influence of relevant major clinical
187 factors on AR development within 90 days after LTx was analyzed (Table 2). The peak IgM
188 level was a significant risk factor with a hazard ratio of 15.642 in univariate analysis ($p=0.001$).
189 In multivariate analyses, the peak value of the IgM rise rate was an independent factor
190 predicting subsequent development of AR. As for the analysis of the impact on CLAD (Table 3),
191 the peak IgM early after LTx exhibited a much higher hazard ratio (3.3) than other clinical

192 variables, although this difference was not statistically significant ($p=0.069$). No other items had
193 evident influence.

194

195 **Analysis for cutoff value determination and comparison between positive and negative**

196 **IgM groups**

197 A receiver operating characteristics curve analysis determined 1.8 as the cutoff
198 anti-HLA IgM level for AR development (Figure 3). Patients were then divided into two groups
199 according to their values. Seven patients had positive anti-HLA IgM levels (1.8 times or higher
200 elevation) and 24 were negative (< 1.8).

201 Transitions of the average IgM levels in the positive and negative groups are shown in
202 Figure 1B. The positive IgM group exhibited a substantially wider fluctuation in the IgM level
203 than the negative IgM group. A comparison between the positive and negative anti-HLA IgM
204 groups is shown in Table 4. Six of 7 patients in the positive group had AR episodes, while only
205 1 of 24 developed AR in the negative group ($p=0.001$). There was a higher proportion of
206 interstitial pneumonia as the primary disease for LTx in the positive group ($p=0.04$). There was
207 no association of PGD grade or other postoperative inflammatory events with anti-HLA IgM
208 elevation.

209

210 **Long term outcomes by anti-HLA IgM levels**

211 The median observation period of the survivors was 63 months (range, 42–88 months).
212 Patients in the positive anti-HLA IgM group developed CLAD more frequently ($p=0.03$)
213 (Figure 4A). Four of 7 patients in the positive group developed CLAD, while 4 of 24 patients in
214 the negative group developed CLAD. As for mortality after LTx, the positive IgM group had
215 poorer survival outcomes than the negative group, although the difference did not reach
216 statistical significance ($p=0.17$) (Figure 4B). Three of the 7 recipients with positive IgM died
217 during the observation period; the cause of death in all the patients was CLAD, one of which
218 developed comorbid reluctant acute AMR episodes.

219

220 **Discussion**

221 Although the immunologically risky pairing of donor and recipient (e.g., evidence of
222 preformed DSA or positive lymphocyte crossmatching test) is usually precluded in the common
223 organ allocation policy, the challenge of management against frequent acute and chronic
224 rejection is still inevitable after LTx. Susceptibility of the transplanted organs to rejection varies
225 by HLA differences between the donor and recipient. However, it is common for every
226 post-transplant management to start and continue immunosuppressant treatment with a
227 standardized dose and concentration according to a generalized protocol. The dosage of

228 immunosuppressants is usually modified and customized after the development of the clinical
229 signs of rejection or drug side effects. This is mainly attributed to the fact that there is no early
230 clinical marker in use to predict how likely rejection develops later in each transplant case. This
231 study focused on anti-HLA IgM production in the recipient early after LTx to address this issue.
232 We explored serum IgM not from an aspect of actual damaging factor but as a prognostic
233 marker in LTx recipients in this study.

234 We identified an increase in the risk of AR, CLAD, and mortality in patients with
235 anti-HLA-IgM surge early after LTx. Patients with more than 1.8-fold (approximately double)
236 increase in anti-HLA IgM during the first two weeks after LTx had higher risks of subsequent
237 graft rejection than those with less than double. The IgM rise was determined by means of
238 paired-serum examination (proportion to baseline level) because patients could originally have
239 natural anti-HLA or cross-reactive IgM, which influenced the baseline values on an individual
240 basis. The starting immunosuppressant management policy was standardized during the study
241 period with equal target trough levels of tacrolimus and steroid tapering policy. De novo
242 HLA-IgM production is largely unaffected by various other inflammatory events such as
243 primary graft dysfunction. Overall, the rise in anti-HLA IgM can be one of the early predictors
244 of subsequent immune reactivity and graft survival.

245 The detailed mechanism of the association between humoral immune response and T

246 cell mediated rejection or chronic rejection is not well known. However, there are considerable
247 clinical evidence previously reported, indicating potentially close correlation among them.
248 According to a study of a large-scale series of kidney transplantations, acute rejection episodes
249 were significantly associated with the development of de novo DSA [17]. Furthermore, other
250 recent studies demonstrated that the combination of T cell and antibody mediated rejection is
251 much frequently observed in the condition of chronic allograft failure rather than purely
252 mechanized rejection [18-20]. Cellular immunity induced by CD4+ Th2 cells can prompt
253 specific B cells to produce DSA during acute rejection [21]. The Th1 and Th2 cells mutually
254 inhibit each other's activity through cytokine signals but can coexist. We previously reported the
255 early elevation of circulating immunoglobulin (Ig) M levels against donor lymphocytes in
256 histologically proven acute cellular rejection in rat allo-LTx models [12, 13]. Those studies
257 suggested that a dynamic humoral immune response occurs even in cellular rejection and links
258 to chronic allograft failure. In addition, it is well recognized that repeated acute rejection early
259 after LTx is associated with higher risk of later development of bronchiolitis obliterans
260 syndrome [22]. The early fluctuation of anti-HLA IgM we observed was a primitive but
261 sensitive change that could represent general immune reactivity against the allograft antigenicity.
262 We consider that monitoring early immune response after transplantation would be a reasonable
263 way to predict and grade the subsequent immunological risk after transplantation.

264 IgM is the first isotype of immunoglobulin that emerges in the blood after initiation of
265 antigen recognition. It is controversial whether donor specific IgM has functionality to injure
266 the graft by itself and the specificity of IgM is equivalent to IgG. However, its serum levels
267 sensitively rise in response to donor antigenicity prior to class switching. IgM against donor
268 HLA was similarly detected in kidney and heart transplant recipients with undermined survival
269 outcomes [23]. As for the characteristics of the major histocompatibility complex (MHC)
270 antigen of the graft, class I is widely expressed on the graft tissues and recognized as an
271 alloantigen very early after transplantation [24,25]. Although de novo antibodies against MHC
272 class II antigens mainly contribute to persistent rejection and graft dysfunction in the long-term
273 [26], upregulation of class II antigen provoking an immunoreactivity can occur after some
274 processes of graft injury. Theoretically, therefore, antibodies toward MHC class II usually
275 emerge later than antibodies toward class I [25]. We did not find significant fluctuations in IgM
276 toward class II antigens during the first 14 days after LTx in a preliminary study (data not
277 shown). Overall, it is reasonable to consider that elevation of IgM levels against class I HLA
278 can be an early sensitive maker of subsequent rejection and graft survival.

279 We employed the flow PRA technique to screen anti-HLA IgM in this study. Although
280 a flow PRA screening kit was originally used for IgG detection and qualitative analysis, the
281 methodology to transfer it to an anti-HLA IgM assay was reported previously [27]. In the

282 framework of the flow PRA test used in this study, it was not proven whether the increased
283 anti-HLA IgM components were truly donor-specific or not. However, the specific antibody to
284 the specific donor HLA alleles is not targetable to quantify as the most effective marker because
285 the types of HLA antigens that provoke early responsive antibody production are unpredictable.
286 In addition, repeated examination to search for the peak levels for specific antibodies (with a
287 single antigen technique) is quite expensive. Therefore, it is a logical and practical way to
288 monitor the whole anti-HLA IgM value by applying the flow PRA method as shown in this
289 study.

290 There are several limitations inherent to the design of our study. It is retrospective and
291 small-scale in nature. Dose management for immunosuppressants after rejection development
292 was not standardized and conducted in the clinical judgment on an individual case basis. Donor
293 specificity of the increased IgM was not validated to simplify the monitoring methodology as
294 mentioned above. The negativity of pretransplant anti-HLA IgG was determined by the
295 conventional complement-dependent cytotoxicity method. The study was focused on the impact
296 of anti-HLA IgM as a very early sensitive maker of rejection rather than its specificity and the
297 effect of de novo IgG was not evaluated here. The qualitative methodology of conventional flow
298 PRA technique was transferred to the way to measure MFI of the overall anti-HLA IgM titer.
299 Furthermore, the diagnosis of AR was made clinically but not histologically. It is necessary to

300 take them into consideration when interpreting the study results.

301 In conclusion, elevation of anti-HLA IgM levels early after LTx potentially
302 correlated with a higher incidence of later rejection and worse outcomes. Monitoring the level
303 of anti-HLA IgM early after transplant can contribute to subsequent optimal
304 immunosuppressive adjustments. However, whether the IgM value can reflect the
305 immunoreactivity similarly in the chronic phase remains an open question. A further study to
306 observe IgM-positive recipients and determine how to manage immunosuppressant
307 management is warranted.

308 **AUTHOR CONTRIBUTIONS**

309 Kazuaki Miyahara, MD: Data collection and analysis, statistics, drafting article

310 Kentaroh Miyoshi, MD, PhD: Clinical practice, concept / design, data interpretation, drafting
311 and revising the article

312 Takeshi Kurosaki MD, PhD: Clinical practice, data collection

313 Shinji Otani MD, PhD: Clinical practice, data collection

314 Seiichiro Sugimoto MD, PhD: Clinical practice, data collection

315 Masaomi Yamane MD, PhD: Clinical practice, data collection, critical revision of the article

316 Shinichi Toyooka MD, PhD: Clinical practice, data interpretation, critical revision of the article

317

318 **Acknowledgements:**

319 This research was supported by a Grant-in-Aid for Scientific Research C (Grant numbers:

320 17K10785 and 20K0917602). The authors declare no conflicts of interest.

321

322 **Conflict of interest statement:**

323 Kazuaki Miyahara and other co-authors have no conflict of interest.

324 **References**

- 325 1. The International Society for Heart and Lung Transplantation. International Thoracic Organ
326 Transplant Registry Data Slides. <https://ishltregistries.org/registries/slides.asp>
- 327 2. Kawashima M, Juvet SC. The role of innate immunity in the long-term outcome of lung
328 transplantation. *Ann Transl Med* 2020;8:412.
- 329 3. Bharat A, Kuo E, Steward N, Aloush A, Hachem R, Trulock EP, et al. Immunological link
330 between primary graft dysfunction and chronic lung allograft rejection. *Ann Thorac Surg*
331 2008;86:189-95
- 332 4. Verleden SE, Von der Thüsen J, Roux A, Brouwers ES, Braubach P, Kuehnel M, et al. When
333 tissue is the issue: A histological review of chronic lung allograft dysfunction. *Am J Transplant*
334 2020;20:2644-2651.
- 335 5. Hachem RR. The role of the immune system in lung transplantation: towards improved
336 long-term results. *J Thorac Dis* 2019;11(Suppl 14):S1721-S1731.
- 337 6. Shtraichman O, Diamond JM. Emerging biomarkers in chronic lung allograft
338 dysfunction. *Expert Rev Mol Diagn* 2020;20:467-475.
- 339 7. Verleden SE, Vanaudenaerde BM, Emonds MP, Van Raemdonck DE, Neyrinck AP,
340 Verleden GM, et al. Donor-specific and -nonspecific HLA antibodies and outcome post lung
341 transplantation. *Eur Respir J* 2017;50:1701248.

- 342 8. Morrell MR, Pilewski JM, Gries CJ, Pipeling MR, Crespo MM, Ensor CR, et al. De novo
343 donor-specific HLA antibodies are associated with early and high-grade bronchiolitis obliterans
344 syndrome and death after lung transplantation. *J Heart Lung Transplant* 2014;33:1288-1294.
- 345 9. Le Pavec J, Suberbielle C, Lamrani L, Feuillet S, Savale L, Dorfmueller P, et al. De-novo
346 donor-specific anti-HLA antibodies 30 days after lung transplantation are associated with a
347 worse outcome. *J Heart Lung Transplant* 2016;35:1067-1077.
- 348 10. Palmer SM, Davis RD, Hadjiliadis D, Hertz MI, Howell DN, Ward FE, et al. Development
349 of an antibody specific to major histocompatibility antigens detectable by flow cytometry after
350 lung transplant is associated with bronchiolitis obliterans syndrome. *Transplantation*
351 2002;74:799-804.
- 352 11. Miyoshi K, Sano Y, Yamane M, Toyooka S, Oto T, Miyoshi S. Elevation of antidonor
353 immunoglobulin M levels precedes acute lung transplant rejection. *Ann Thorac Surg*
354 2011;92:1233-8.
- 355 12. Yamane M, Sano Y, Shimizu N. Significant changes in the alloantibody after lung
356 transplantation in the cyclosporine treated rat model. *Transpl Immunol* 2004;12:143-150.
- 357 13. Yamane M, Sano Y, Nagahiro I, Aoe M, Date H, Ando A, et al. Humoral immune responses
358 during acute rejection in rat lung transplantation. *Transpl Immunol* 2003;11:31-37.

- 359 14. Hoshikawa Y, Okada Y, Ashikari J, Matsuda Y, Niikawa H, Noda M, et al. Medical
360 consultant system for improving lung transplantation opportunities and outcomes in
361 Japan. *Transplant Proc* 2015;47:746-750.
- 362 15. Verleden GM, Glanville AR, Lease ED, Fisher AJ, Calabrese F, Corris PA, et al. Chronic
363 lung allograft dysfunction: Definition, diagnostic criteria, and approaches to treatment - A
364 consensus report from the Pulmonary Council of the ISHLT. *J Heart Lung Transplant*
365 2019;38:493-503.
- 366 16. Christie JD, Carby M, Bag R, Corris P, Hertz M, Weill D; ISHLT Working Group on
367 Primary Lung Graft Dysfunction. Report of the ISHLT Working Group on Primary Lung Graft
368 Dysfunction part II: definition. A consensus statement of the International Society for Heart and
369 Lung Transplantation. *J Heart Lung Transplant* 2005;24:1454-1459.
- 370 17. Hourmant M, Cesbron-Gautier A, Terasaki PI, Mizutani K, Moreau A, Meurette A, et al.
371 Frequency and clinical implications of development of donor-specific and non-donor-specific
372 HLA antibodies after kidney transplantation. *J Am Soc Nephrol.* 2005;16:2804-2812.
- 373 18. Wiebe C, Gibson IW, Blydt-Hansen TD, Pochinco D, Birk PE, Ho J, et al. Rates and
374 determinants of progression to graft failure in kidney allograft recipients with de novo
375 donor-specific antibody. *Am J Transplant.* 2015;15:2921-2930.
- 376 19. Cherukuri A, Mehta R, Sharma A, Sood P, Zeevi A, Tevar AD, Rothstein DM, Hariharan S.

377 Post-transplant donor specific antibody is associated with poor kidney transplant outcomes only
378 when combined with both T-cell-mediated rejection and non-adherence. *Kidney Int.*
379 2019;96:202-213.

380 20. Chong AS. Mechanisms of organ transplant injury mediated by B cells and antibodies:
381 Implications for antibody-mediated rejection. *Am J Transplant.* 2020;20 Suppl 4:23-32.

382 21. Steele DJ, Laufer TM, Smiley ST, Ando Y, Grusby MJ, Glimcher LH, et al. Two levels of
383 help for B cell alloantibody production. *J Exp Med.* 1996;183:699-703.

384 22. Meyer KC, Raghu G, Verleden GM, Corris PA, Aurora P, Wilson KC, et al;
385 ISHLT/ATS/ERS BOS Task Force Committee; ISHLT/ATS/ERS BOS Task Force Committee.
386 An international ISHLT/ATS/ERS clinical practice guideline: diagnosis and management of
387 bronchiolitis obliterans syndrome. *Eur Respir J.* 2014;44:1479-503.

388 23. Stastny P, Ring S, Lu C, Arenas J, Han M, Lavingia B. Role of immunoglobulin (Ig)-G and
389 IgM antibodies against donor human leukocyte antigens in organ transplant recipients. *Hum*
390 *Immunol* 2009;70:600-604.

391 24. Gabriel C, Fürst D, Faé I, Wenda S, Zollikofer C, Mytilineos J, et al. HLA typing by
392 next-generation sequencing - getting closer to reality. *Tissue Antigens* 2014;83:65-75.

393 25. Nakamura T, Shirouzu T, Nakata K, Yoshimura N, Ushigome H. The role of major
394 histocompatibility complex in organ transplantation- Donor specific anti-major

395 histocompatibility complex antibodies analysis goes to the next stage. *Int J Mol Sci*
396 2019;20:4544.

397 26. Koutsokera A, Royer PJ, Antonietti JP, Fritz A, Benden C, Aubert JD, et al. Development of
398 a multivariate prediction model for early-onset bronchiolitis obliterans syndrome and restrictive
399 allograft syndrome in lung transplantation. *Front Med (Lausanne)* 2017;4:109.

400 27. Khan N, Robson AJ, Worthington JE, Martin S. The detection and definition of IgM
401 alloantibodies in the presence of IgM autoantibodies using flow PRA beads. *Hum Immunol*
402 2003;64:593-599.

403 **Legends**

404 Figure 1. Transition of average anti-HLA IgM level in the overall cohort (A), negative and
405 positive-IgM groups (B). The IgM levels are calculated as rise rates of MFI compared to the
406 baseline of the day 2 MFI level in each case.

407 Figure 2. A monitoring of anti-HLA IgM in seven patients with acute rejection episodes. Steroid
408 pulse therapy was initiated on the day of AR diagnosis. Six of the seven patients (except case 2)
409 exhibited a clear surge of anti-HLA IgM level surpassing 1.8 of rise rate in the first 14-day
410 monitoring period. The IgM levels were determined by means of paired MFI examination where
411 baseline was defined as MFI value of day 2.

412 Figure 3. Receiver operating characteristic curve indicating a cutoff level of anti-HLA IgM for
413 prediction of subsequent acute rejection. Optimal sensitivity and specificity were achieved when
414 the cutoff value was set at 1.8.

415 Figure 4. A Kaplan-Meier analysis for the chronic lung allograft dysfunction free survival (A)
416 and overall survival (B). The positive and negative IgM groups are compared.