

Review

## Histidine-rich Glycoprotein Modulates the Blood-vascular System in Septic Condition

Hidenori Wake\*§

*Department of Pharmacology, Okayama University Graduate School of Medicine, Dentistry,  
and Pharmaceutical Sciences, Okayama 700-8558, Japan*

Histidine-rich glycoprotein (HRG) is a 75 kDa glycoprotein synthesized in the liver whose plasma concentration is 100-150 µg/ml. HRG has been shown to modulate sepsis-related biological reactions by binding to several substances and cells, including heparin, factor XII, fibrinogen, thrombospondin, plasminogen, C1q, IgG, heme, LPS, dead cells, bacteria, and fungi. Therefore, reduction of plasma HRG levels in sepsis leads to dysregulation of coagulation, fibrinolysis, and immune response, resulting in disseminated intravascular coagulation and multiple organ failure. This review summarizes the binding and functional properties of HRG in sepsis.

**Key words:** histidine-rich glycoprotein, septic pathogenesis, immunothrombosis

**H**istidine-rich glycoprotein (HRG), first isolated as histidine-rich 3.8S α<sub>2</sub>-glycoprotein in 1972 [1,2], is a 75 kDa plasma glycoprotein mainly produced by the liver at approximately 100-150 µg/ml in human blood [3,4]. It is composed of 4 domains: cystatin-like domain 1; cystatin-like domain 2; histidine-proline rich domain, which has a unique 12 GHHPH tandem repeat amino acid sequence; and a C-terminal domain [5,6]. HRG is present in the blood of vertebrates such as humans, rats, mice, rabbits, and chickens, and a few invertebrates [3,6,7]. HRG can bind a variety of ligands such as heparin, factor XII (FXII), fibrinogen, thrombospondin, plasminogen, C1q, IgG, heme, and lipopolysaccharide (LPS), and can interact with dead cells, bacteria, and fungi. These binding properties indicate that HRG modulates coagulation [8-14] and fibrinolysis [15-18], helps to scavenge toxic substances [19-25] and dead cells [26-28], and kills

pathogens [29-32]. In addition, our recent studies have reported that the plasma concentration of HRG was strikingly reduced during severe systemic inflammatory and septic condition triggered by infection, leading to immunothrombosis and tissue damage, and subsequent disseminated intravascular coagulation (DIC) and multiple organ failure (MOF) (Fig.1) [33-40]. These results suggest that HRG is involved in the regulation of coagulation and fibrinolysis and the immune system in septic condition. This review is focused on the function of HRG in these systems.

### Functions of HRG in Coagulation and Fibrinolysis

Coagulation and fibrinolysis are rigorously regulated by several biological substances to prevent blood loss and unnecessary thrombus formation. HRG is reported to bind to coagulation and fibrinolysis-related molecules such as heparin, FXII, fibrinogen, thrombospondin, and plasminogen [8-18].

Received February 28, 2019; accepted April 26, 2019.

\*Corresponding author. Phone & Fax : +81-86-235-7138

E-mail : wake-h@cc.okayama-u.ac.jp (H. Wake)

§The winner of the 2017 Incentive Award of the Okayama Medical Association in General Medical Science.

Conflict of Interest Disclosures: No potential conflict of interest relevant to this article was reported.

**Heparin.** The anti-coagulant activity of anti-thrombin III (ATIII) is enhanced in the presence of heparin. HRG binds to heparin with high affinity, blocking the interaction between heparin and ATIII, and inhibiting its anti-coagulant activity [8-10].

**FXII.** HRG also modulates the intrinsic pathway through high-affinity binding to FXII, inhibiting FXII autoactivation and FXIIa-mediated activation of FXI [11].

**Fibrinogen and thrombospondin.** Although HRG interaction with fibrinogen, the platelet-bridging molecule, and subsequent incorporation into a fibrin clot does not induce the conversion of fibrinogen to fibrin within the clot, this interaction influences the structure of the fibrin gel [12]. HRG inhibits the interaction between GPIIb/IIIa and fibrinogen via its histidine-rich domain in a  $Zn^{2+}$ -dependent manner and inactivates the platelet-aggregation promoter thrombospondin on the surface of platelets, thereby interfering with platelet-platelet interactions [13, 14].

**Plasminogen.** HRG inhibits the interaction of plasminogen with fibrin/fibrinogen and retards fibrinolysis due to its binding to the plasminogen lysine-binding site competing with fibrin binding [15]. Contrarily, immobilized HRG on cell surface heparan sulfate results in 30-fold increase in the conversion of plasminogen to plasmin by the fibrinolysis promoter tPA [16-18].

**Immunothrombosis.** Immunothrombosis is a key phenomenon in the development of septic pathogenesis, which is triggered by activated-neutrophil adhesion to vascular endothelial cells, and subsequent platelet accumulation and fibrin polymer deposition on their surface. Immunothrombosis helps prevent the diffusion

of pathogens to the systemic blood stream and their disposal in physiological conditions. However, impairment of this regulation in sepsis causes immunothrombus formation to lead to DIC and subsequent MOF [37-40]. Our recent study revealed that HRG inhibits immunothrombus formation by keeping circulating neutrophils quiescent (Fig. 1) [35, 36].

### Functions of HRG in Immunity and Inflammation

**Insoluble immune complexes (IICs).** Although immune complexes promote the clearance of pathogens and foreign substances, when IICs form and deposit in several organs, they lead to tissue injury and inflammation. C1q enhances IIC formation by directly binding and inducing the cross-linking of IgG molecules [19]. The N-terminal domain of HRG binds to C1q and the F(ab) region of IgG, inhibiting the formation of IICs composed of ovalbumin and the anti-ovalbumin antibody [20]. This domain binding also inhibits IIC formation by preventing Fc-Fc interactions by masking IgG epitopes recognized by rheumatoid factor. Furthermore, HRG promotes the solubilization and clearance of IICs [21].

**Heme and LPS.** Heme and LPS are damage-associated molecular pattern (DAMP)/pathogen-associated molecular pattern (PAMP) molecules, respectively, that promote tissue injury. HRG binds heme, neutralizing heme cytotoxicity and inhibiting hemolysis [22-24]. Synthetic peptides derived from the histidine-rich domain of HRG also bind and neutralize LPS, inhibiting LPS-stimulated IL-8 production of CD14-transfected THP-1 cells [25].

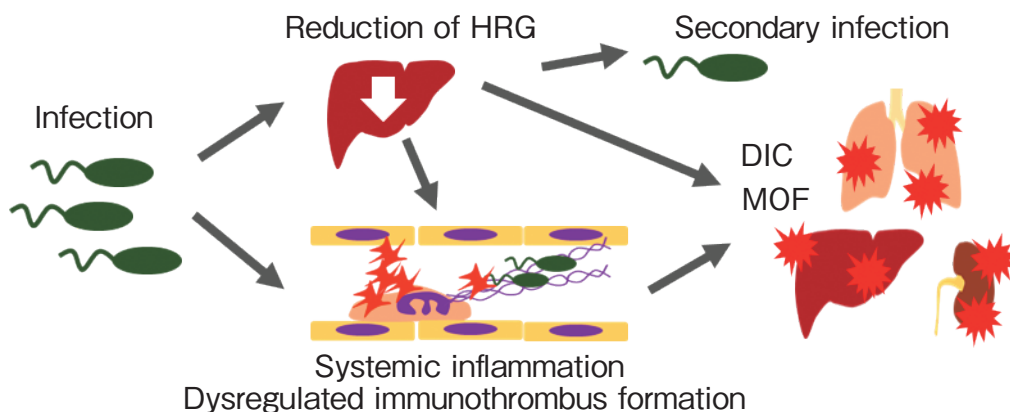


Fig. 1 Scheme of septic pathogenesis. DIC, disseminated intravascular coagulation; MOF, multiple organ failure.

**Dying and dead cells.** Rapid removal of apoptotic and necrotic cells from blood stream and tissue by phagocytes is necessary to maintain tissue homeostasis. Impairing clearance of these cells leads to the release of DAMP molecules, which induce inflammation and tissue damage. HRG can aid in the phagocytosis of apoptotic cells by macrophages, by acting as a bridge between FcγRI on macrophages and DNA on the surface of apoptotic cells [26]. Additionally, HRG recognizes intracellular phospholipids exposed in necrotic cells and aids in the recruitment of IgG, leading to the phagocytosis of necrotic cells via FcγRI and heparan sulfate on phagocytes [27, 28].

**Antimicrobial activity.** Moreover, HRG exerts antimicrobial activity by binding to the surface of the Gram-positive bacteria *Enterococcus faecalis* and *Streptococcus pyogenes*, the Gram-negative bacteria *Escherichia coli*, and the fungus *Candida albicans* under acidic conditions or in the presence of Zn<sup>2+</sup> and inducing membrane destabilization [29–32].

## Concluding Remarks

Septic pathogenesis can result from systemic coagulation, fibrinolysis, and inflammation. This review revealed that HRG can ameliorate septic condition by modulating these biological reactions through high-affinity binding to several key molecules and cells (Fig. 1). Thus, HRG has potential value as a therapeutic drug for the treatment of sepsis.

**Acknowledgments.** This work was supported by grant from the Japan Agency for Medical Research and Development, AMED (18lk0201085h0001). We would like to thank Editage (www.editage.jp) for English language editing.

## References

- Haupt H and Heimbürger N: Human serum proteins with high affinity for carboxymethylcellulose. I. Isolation of lysozyme, C1q and 2 hitherto unknown globulins. *Hoppe Seylers Z Physiol Chem* (1972) 353: 1125–1132.
- Heimbürger N, Haupt H, Kranz T and Baudner S: Human serum proteins with high affinity to carboxymethylcellulose. II. Physicochemical and immunological characterization of a histidine-rich 3,8S-2-glycoprotein (CM-protein I). *Hoppe Seylers Z Physiol Chem* (1972) 353: 1133–1140.
- Corrigan JJ, Jeter MA, Bruck D and Feinberg WM: Histidine-rich glycoprotein levels in children: the effect of age. *Thromb Res* (1990) 59: 681–686.
- Saito H, Goodnough LT, Boyle JM and Heimbürger N: Reduced histidine-rich glycoprotein levels in plasma of patients with advanced liver cirrhosis. Possible implications for enhanced fibrinolysis. *Am J Med* (1982) 73: 179–182.
- Jones AL, Hulett MD and Parish CR: Histidine-rich glycoprotein: a novel adaptor protein in plasma that modulates the immune, vascular and coagulation systems. *Immunol Cell Biol* (2005) 83: 106–118.
- Koide T, Foster D, Yoshitake S and Davie EW: Amino acid sequence of human histidine-rich glycoprotein derived from the nucleotide sequence of its cDNA. *Biochemistry* (1986) 25: 2220–2225.
- Drasin T and Sahud M: Blood-type and age affect human plasma levels of histidine-rich glycoprotein in a large population. *Thromb Res* (1996) 84: 179–188.
- Koide T, Odani S and Ono T: The N-terminal sequence of human plasma histidine-rich glycoprotein homologous to antithrombin with high affinity for heparin. *FEBS Lett* (1982) 141: 222–224.
- Lijnen HR, van Hoef B and Collen D: Interaction of heparin with histidine-rich glycoprotein and with antithrombin III. *Thromb Haemost* (1983) 50: 560–562.
- Lijnen HR, Van Hoef B and Collen D: Histidine-rich glycoprotein modulates the anticoagulant activity of heparin in human plasma. *Thromb Haemost* (1984) 51: 266–268.
- MacQuarrie JL, Stafford AR, Yau JW, Leslie BA, Vu TT, Fredenburgh JC and Weitz JI: Histidine-rich glycoprotein binds factor XIIa with high affinity and inhibits contact-initiated coagulation. *Blood* (2011) 117: 4134–4141.
- Leung LL: Interaction of histidine-rich glycoprotein with fibrinogen and fibrin. *J Clin Invest* (1986) 77: 1305–1311.
- Leung LL, Nachman RL and Harpel PC: Complex formation of platelet thrombospondin with histidine-rich glycoprotein. *J Clin Invest* (1984) 73: 5–12.
- Wakabayashi S and Koide T: Histidine-rich glycoprotein: a possible modulator of coagulation and fibrinolysis. *Semin Thromb Hemost* (2011) 37: 389–394.
- Lijnen HR, Hoylaerts M and Collen D: Isolation and characterization of a human plasma protein with affinity for the lysine binding sites in plasminogen. Role in the regulation of fibrinolysis and identification as histidine-rich glycoprotein. *J Biol Chem* (1980) 255: 10214–10222.
- Borza D-B and Morgan WT: Acceleration of plasminogen activation by tissue plasminogen activator on surface-bound histidine proline-rich glycoprotein. *J Biol Chem* (1997) 272: 5718–5726.
- Jones AL, Hulett MD, Altin JG, Hogg P and Parish CR: Plasminogen is tethered with high affinity to the cell surface by the plasma protein, histidine-rich glycoprotein. *J Biol Chem* (2004) 279: 38267–38276.
- Silverstein RL, Nachman RL, Leung LL and Harpel PC: Activation of immobilized plasminogen by tissue activator. Multimolecular complex formation. *J Biol Chem* (1985) 260: 10346–10352.
- Gorgani NN, Parish CR, Easterbrook Smith SB and Altin JG: Histidine-rich glycoprotein binds to human IgG and C1q and inhibits the formation of insoluble immune complexes. *Biochemistry* (1997) 36: 6653–6662.
- Gorgani NN, Easterbrook-Smith SB and Altin JG: The formation of insoluble immune complexes between ovalbumin and anti-ovalbumin IgG occurs in at least two distinct phases dependent on reactant concentration and ionic strength. *Biochim Biophys Acta* (1996) 1317: 45–54.
- Gorgani NN, Altin JG and Parish CR: Histidine-rich glycoprotein prevents the formation of insoluble immune complexes by rheumatoid factor. *Immunology* (1999) 98: 456–463.

22. Katagiri M, Tsutsui K, Yamano T, Shimonishi Y and Ishibashi F: Interaction of heme with a synthetic peptide mimicking the putative heme-binding site of histidine-rich glycoprotein. *Biochem Biophys Res Commun* (1987) 149: 1070–1076.
23. Morgan WT: Human serum histidine-rich glycoprotein. I. Interactions with heme, metal ions and organic ligands. *Biochim Biophys Acta* (1978) 535: 319–333.
24. Zhong H, Wake H, Liu K, Gao Y, Teshigawara K, Sakaguchi M, Mori S and Nishibori M: Effects of Histidine-rich glycoprotein on erythrocyte aggregation and hemolysis: Implications for a role under septic conditions. *J Pharmacol Sci* (2018) 136: 97–106.
25. Bosshart H and Heinzelmann M: Endotoxin-neutralizing effects of histidine-rich peptides. *FEBS Lett* (2003) 553: 135–140.
26. Gorgani NN, Smith BA, Kono DH and Theofilopoulos AN: Histidine-rich glycoprotein binds to DNA and Fc gamma RI and potentiates the ingestion of apoptotic cells by macrophages. *J Immunol* (2002) 169: 4745–4751.
27. Poon IK, Hulett MD and Parish CR: Histidine-rich glycoprotein is a novel plasma pattern recognition molecule that recruits IgG to facilitate necrotic cell clearance *via* FcγRI on phagocytes. *Blood* (2010) 115: 2473–2482.
28. Poon IK, Parish CR and Hulett MD: Histidine-rich glycoprotein functions cooperatively with cell surface heparan sulfate on phagocytes to promote necrotic cell uptake. *J Leukoc Biol* (2010) 88: 559–569.
29. Kacprzyk L, Rydengård V, Mörgelin M, Davoudi M, Pasupuleti M, Malmsten M and Schmidtchen A: Antimicrobial activity of histidine-rich peptides is dependent on acidic conditions. *Biochim Biophys Acta* (2007) 1768: 2667–2680.
30. Rydengård V, Olsson A-K, Mörgelin M and Schmidtchen A: Histidine-rich glycoprotein exerts antibacterial activity. *FEBS J* (2007) 274: 377–389.
31. Rydengård V, Shannon O, Lundqvist K, Kacprzyk L, Chalupka A, Olsson AK, Mörgelin M, Jähnen-Dechent W, Malmsten M and Schmidtchen A: Histidine-rich glycoprotein protects from systemic *Candida* infection. *PLoS Pathog* (2008) 4: e1000116.
32. Shannon O, Rydengård V, Schmidtchen A, Mörgelin M, Alm P, Sørensen OE and Björck L: Histidine-rich glycoprotein promotes bacterial entrapment in clots and decreases mortality in a mouse model of sepsis. *Blood* (2010) 116: 2365–2372.
33. Kuroda K, Wake H, Mori S, Hinotsu S, Nishibori M and Morimatsu H: Decrease in histidine-rich glycoprotein as a novel biomarker to predict sepsis among systemic inflammatory response syndrome. *Crit Care Med* (2018) 46: 570–576.
34. Saigo K, Yoshida A, Ryo R, Yamaguchi N and Leung LL: Histidine-rich glycoprotein as a negative acute phase reactant. *Am J Hematol* (1990) 34: 149–150.
35. Terao K, Wake H, Adachi N, Liu K, Teshigawara K, Takahashi H, Mori S and Nishibori M: Histidine-rich glycoprotein suppresses hyperinflammatory responses of lung in a severe acute pancreatitis mouse model. *Pancreas* (2018) 47: 1156–1164.
36. Wake H, Mori S, Liu K, Morioka Y, Teshigawara K, Sakaguchi M, Kuroda K, Gao Y, Takahashi H, Ohtsuka A, Yoshino T, Morimatsu H and Nishibori M: Histidine-rich glycoprotein prevents septic lethality through regulation of immunothrombosis and inflammation. *EBioMedicine* (2016) 9: 180–194.
37. Seymour CW, Liu VX, Iwashyna TJ, Brunkhorst FM, Rea TD, Scherag A, Rubenfeld G, Kahn JM, Shankar-Hari M, Singer M, Deutschman CS, Escobar GJ and Angus DC: Assessment of clinical criteria for sepsis: for the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* (2016) 315: 762–774.
38. Delabranche X, Helms J and Meziani F: Immunohaemostasis: a new view on haemostasis during sepsis. *Ann Intensive Care* (2017) 7: 117.
39. Frantzeskaki F, Armaganidis A and Orfanos SE: Immunothrombosis in acute respiratory distress syndrome: cross talks between inflammation and coagulation. *Respiration* (2017) 93: 212–225.
40. Iba T and Levy JH: Inflammation and thrombosis: roles of neutrophils, platelets and endothelial cells and their interactions in thrombus formation during sepsis. *J Thromb Haemost* (2018) 16: 231–241.