

1 *Original Article (Experimental Original)*

2 **Single nucleotide polymorphism in a gene modulating glucocorticoid sensitivity**
3 **is associated with decline in total lung capacity after lung transplantation**

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22 lung capacity; Chronic lung allograft dysfunction

1 **Abstract**

2 **Purpose:** Glucocorticoids are used to prevent chronic lung allograft dysfunction (CLAD)
3 after lung transplantation (LT). Our study was aimed at assessing the association
4 between the glucocorticoid-induced transcript 1 gene (GLCCI1) variant, which
5 modulates glucocorticoid sensitivity, and postoperative lung function and the
6 development of CLAD after LT.

7 **Methods:** A total of 71 recipients of LT were genotyped for the GLCCI1 variant,
8 rs37972, and divided into 3 groups: the homozygous mutant allele (TT) group, the
9 heterozygous mutant allele (CT) group, and the wild-type allele (CC) group. The results
10 of pulmonary function tests were compared with the postoperative baseline values.

11 **Results:** The total lung capacity (TLC) in the TT group was significantly lower than that
12 in the CC group at 3 years after LT ($P = 0.029$). In the recipients of cadaveric LT, the
13 TLC and forced expiratory volume in 1 second in the TT group were significantly lower
14 than those in the CC groups, resulting in a significant worse CLAD-free survival at 3
15 years after LT ($P = 0.016$).

16 **Conclusion:** The GLCCI1 variant was associated with a significant decrease of the TLC
17 at 3 years after LT and the development of CLAD at 3 years, especially in patients
18 undergoing cadaveric LT. Genotyping for this SNP might prove useful for adjusting the
19 glucocorticoid dose to prevent a decline of the TLC after LT and reduce the risk of CLAD
20 after cadaveric LT.

21

1 **Introduction**

2 Long-term survival after lung transplantation (LT) still remains worse than that after
3 other solid organ transplantations [1-3], and chronic lung allograft dysfunction (CLAD) is
4 a major obstacle to long-term survival after LT. To prevent CLAD after LT, adequate
5 immunosuppression is key [4, 5]. Currently, the most commonly used maintenance
6 immunosuppression regimen after LT is a triple regimen consisting of a calcineurin
7 inhibitor, mycophenolate mofetil and a glucocorticoid [3]. Of these, the doses of the
8 calcineurin inhibitor and mycophenolate mofetil are currently adjusted by monitoring the
9 trough blood levels of the drug in each patient, whereas the dose of the glucocorticoid is
10 uniformly tapered to 5 to 10 mg per day for maintenance therapy, according to the body
11 weight or body surface area of the patients. Because individual glucocorticoid sensitivity
12 is known to be influenced by various factors [6-11], monitoring of the therapeutic levels
13 of glucocorticoids is not a realistic option. Nonetheless, recipients of LT generally
14 require glucocorticoids over their lifetimes, although successful withdrawal of
15 glucocorticoids after LT has previously been reported in a few patients after LT [12, 13].
16 Owing to the difficulty in monitoring the therapeutic levels of glucocorticoids, the
17 routinely used maintenance dose of glucocorticoids might be insufficient for patients
18 with poor glucocorticoid sensitivity, which could influence the risk of onset of CLAD after
19 LT.

20 Recently, a screening analysis of more than 530000 SNPs revealed that a
21 functional single nucleotide polymorphism (SNP), rs37972, which maps to the
22 glucocorticoid-induced transcript 1 gene (GLCCI1) and modulates glucocorticoid
23 sensitivity has been shown to be associated with a decreased efficiency of

1 glucocorticoid inhalation therapy in patients with asthma and chronic obstructive
2 pulmonary disease (COPD) [14, 15]. Because the GLCCI1 gene is strongly expressed
3 in the lung and lymphoid tissues, including the T and B lymphocytes, the pharmacologic
4 response to glucocorticoids and the postoperative lung graft function in recipients of LT
5 might also be influenced by the presence of this functional GLCCI1 variant, similar to
6 the case in asthma and COPD mentioned above [14, 15]. However, the association
7 between presence of the functional GLCCI1 variant with the postoperative lung graft
8 function in recipients of LT remains to be clearly elucidated.

9 In this study, we investigated the association between the functional GLCCI1 gene
10 polymorphism modulating glucocorticoid sensitivity and postoperative lung graft
11 function, including the risk of development of CLAD after LT.

12

13 **Methods**

14 **Patients**

15 This was a single-center retrospective cohort study conducted in a cohort of 71
16 patients who underwent LT, including 33 cases of cadaveric LT (CLT) and 38 cases of
17 living-donor lobar lung transplantation (LDLLT), at Okayama University Hospital
18 between October 1998 and June 2014. Blood samples were collected from all 71
19 patients between September 2016 and August 2017. The maximum total number of
20 human leukocyte antigen (HLA) mismatches could equal 12 in bilateral LDLLT involving
21 2 different donors, and 6 in single LDLLT and CLT involving only one donor. The study
22 protocol (No.1610-037) was approved by the institutional review board of Okayama
23 University Hospital. Written informed consent for the study was obtained from each of

1 the patients. All methods were performed in accordance with the relevant guidelines and
2 regulations.

3

4 **SNP genotyping**

5 Genotype analysis for the functional polymorphism of the GLCCI1 gene
6 (rs37972) was conducted in all 71 patients. Genomic DNA was isolated from whole
7 blood with a TaqMan® Sample-to-SNP™ kit (Applied Biosystems, Foster City, CA,
8 USA). Samples were analyzed by a TaqMan genotyping assay using the StepOne™
9 real-time polymerase chain reaction (PCR) system (Applied Biosystems), in a 96-well
10 array plate that included four blank wells as negative controls. The PCR profile
11 consisted of an initial denaturation step at 95°C for 20 secs, 40 cycles at 95°C for 3
12 secs, and at 60°C for 20 secs. The PCR products were analyzed by the StepOne™
13 Software Ver 2.3 (Applied Biosystems). To assess the quality of the genotyping, repeat
14 genotyping was conducted in a randomly selected 5% of the samples, and 100%
15 agreement was confirmed. According to the results of the genotyping, the patients were
16 divided into 3 groups; the homozygous mutant T allele (TT) group, the heterogeneous
17 mutant allele (CT) group, and the homozygous wild-type allele (CC) group.

18

19 **Evaluation of pulmonary function**

20 To evaluate the postoperative changes of the pulmonary function parameters
21 over time, forced expiratory volume in 1 second (FEV1), an indicator to diagnose
22 obstructive CLAD (bronchiolitis obliterans syndrome, BOS) [16], forced vital capacity
23 (FVC), an indicator to diagnose restrictive CLAD (restrictive allograft syndrome, RAS)

1 [16], and total lung capacity (TLC), the originally used indicator to diagnose RAS [17],
2 were measured at 3, 6, 12, 24 and 36 months after LT, and compared with the
3 postoperative stable baseline values among the 3 groups. The postoperative stable
4 baseline values were calculated as the mean values between the two best points in
5 postoperative pulmonary function tests. Additionally, subgroup analysis was performed
6 based on the type of lung donor, cadaveric or living. CLAD was diagnosed using the
7 classification system proposed by the International Society for Heart and Lung
8 Transplantation (ISHLT) [16]. For the differential diagnosis of CLAD, blood examination,
9 chest X-ray, computed tomography of the chest, ventilation-perfusion scanning, the 6-
10 minute walk test, electrocardiogram, and echocardiogram were also performed at the
11 same time as the pulmonary function testing.

12

13 **Statistical analysis**

14 All statistical analyses were performed using the GraphPad Prism 7.04 software
15 (San Diego, CA, USA). The postoperative changes in the pulmonary function
16 parameters were expressed as the mean percentages \pm standard errors of the baseline
17 values. Difference in the baseline characteristics among the groups were tested by the
18 Kruskal-Wallis test for continuous variables, and Pearson's chi-square test for
19 categorical variables. Differences among the % baseline values of the individual
20 measures in the different genotype groups were analyzed by one-way ANOVA, followed
21 by Tukey's test for individual between-group comparisons. The CLAD-free survival rate
22 was analyzed using the Kaplan–Meier method, and the log rank test was used for

1 statistical comparison of the differences among the groups. Differences were
2 considered significant at $P < 0.05$.

3

4 **Results**

5 The patient characteristics are shown in Table 1. The frequencies of GLCCI1
6 gene polymorphism in our cohort was similar to those reported in a previous study [18].
7 The clinical characteristics of the patients were similar among the three groups. There
8 were no significant differences in the previously described risk factors for CLAD,
9 including HLA mismatches, cytomegalovirus mismatches, primary graft dysfunction,
10 acute rejection, and gastroesophageal reflux disease [19]. Total amount of
11 glucocorticoids administered during the first 90 days after LT and the maintenance dose
12 of glucocorticoids did not differ significantly among the three groups. CLAD developed
13 in 9 patients during the first 3 years after LT.

14 As shown in Fig. 1, the percent baseline value of the TLC at 3 years after LT was
15 significantly lower in the TT group as compared to the CC group ($P = 0.029$) (Fig. 1a).
16 By contrast, there were no significant differences in the percent baseline values of the
17 FEV1 or FVC at 3 years after LT among the three genotype groups (Fig. 1b, c). Of note,
18 in the recipients of CLT (Table 2), the percent baseline value of TLC as well as that of
19 FEV1 at 3 years after LT was significantly lower in the TT group than in the CC group
20 (TLC, $P = 0.031$; FEV1, $P = 0.0074$) (Fig. 2a, b).

21 The CLAD-free survival rates at 3 years after LT were similar among the 3 groups
22 (Fig. 3a). However, in the recipients of CLT, the CLAD-free survival rate at 3 years after
23 LT was significantly worse in the TT group than in the CC group (Fig. 3b, $P = 0.016$),

1 whereas no significant differences in the CLAD-free survival rates at 3 years after LT were
2 among the three groups ($P = 0.82$) in the recipients of LDLLT.

3

4 **Discussion**

5 In this study, we found that presence of the GLCCI1 gene polymorphism, rs37972,
6 which modulates glucocorticoid sensitivity, was associated with a significant decrease in
7 the TLC at 3 years after LT, after both CLT and LDLLT. In the recipients of CLT, the
8 functional GLCCI1 variant was associated with a significant decrease of not only the
9 TLC, but also of the FEV1, and also a significantly worse CLAD-free survival rate at 3
10 years after CLT. These results suggest that this SNP might be a possible risk factor for
11 decline of the TLC after LT as well as for decline of the FEV1 and the development of
12 CLAD after CLT.

13 Although glucocorticoid sensitivity in LT recipients has received little attention to
14 date, the recipients with the TT genotype of GLCCI1 showed a lower TLC at 3 years
15 after LT in this study. This result indicates that for patients with a decreased
16 glucocorticoid sensitivity, the routinely used maintenance dose of glucocorticoid might
17 not be sufficient to prevent the decline of the TLC after LT. Individual glucocorticoid
18 sensitivities could be influenced by various factors, including genetic factors, number of
19 cellular glucocorticoid receptors and the availability of glucocorticoids [6-11]. The
20 GLCCI1 genotype may influence dexamethasone-induced apoptosis of immune cells
21 [14, 20]. Moreover, different foci of lymphoid neogenesis in different anatomical
22 compartments of the lung, such as small airways and many anatomical compartments
23 of the lung, could contribute to different phenotypes of CLAD (i.e. BOS and RAS) [21].

1 Therefore, we speculated that in LT recipients with decreased glucocorticoid sensitivity,
2 decreased induction of apoptosis of lymphocytes might contribute to triggering of
3 lymphoid neogenesis, predominantly in the anatomical compartments of the lung,
4 causing decline of the TLC after LT. In addition, our study focused on the GLCCI1
5 variant in the recipients, but not donors. This is because lymphoid neogenesis in the
6 transplanted lung has been shown to be derived from the recipient's bone marrow [22],
7 which is closely involved in the development of allograft rejection after LT. Further
8 examination is required to elucidate the mechanism of decline of the TLC in LT
9 recipients with decreased glucocorticoid sensitivity.

10 With regard to the pulmonary function markers of CLAD, the GLCCI1 variant was
11 associated with a significant difference only in the TLC, but not in the FEV1 or FVC after
12 LT, although in the subgroup analysis, there was a significant difference in the TLC and
13 FEV1 in recipients of CLT. Originally, RAS was characterized by upper lobe-
14 predominant fibrotic changes and a restrictive pulmonary function test profile, defined as
15 $FEV1 \leq 80\%$ of the baseline FEV1, and $TLC \leq 90\%$ of the baseline TLC for ≥ 3 weeks
16 [17]. Instead of TLC, FVC has been proposed as a more practically measured marker to
17 diagnose RAS [16], because TLC is not routinely measured at LT centers worldwide.
18 However, our results suggest that TLC might be more sensitive indicator to diagnose
19 RAS than FVC, as previously described [17].

20 The differences in the outcomes between CLT and LDLLT could reflect the
21 postoperative expansion of the undersized lobar grafts after LDLLT. LDLLT is still a
22 realistic option to resolve severe donor shortage in Japan, and in LDLLT, the right and
23 left lower lobes from two healthy donors are implanted in the recipient in place of the

1 whole lungs. Because the undersized lobar grafts attempt to fit into the large chest
2 cavity of the recipient, the FEV1 and FVC could increase up to 2 years after LDLLT [23].
3 Therefore, after LDLLT, the expansion of the transplanted lobes during the first 2 years
4 could be offset by a decrease of the TLC. Furthermore, because of the immunogenic
5 heterogeneity of the transplanted lobes from two different donors, CLAD develops
6 unilaterally in most recipients after LDLLT, and the unaffected contralateral lobar lung
7 acts as a reservoir [24]. These characteristics might enable maintenance of pulmonary
8 function after LDLLT, and lend support to our finding of the GLCCI1 variant being
9 associated with a worse CLAD-free survival only after CLT. A better understanding of
10 the effect of the GLCCI1 variant in LDLLT recipients might be obtained from long-term
11 follow-up of the patients.

12 Our study had several limitations. First, it was retrospective study conducted at a
13 single transplant center, and the number of patients was small. The follow-up period
14 was still intermediate, and longer follow-up periods would be required for further
15 validation of the incidence of CLAD. This study targeted only Japanese patients,
16 thereby limiting the generalizability of the results.

17 In conclusion, presence of a functional GLCCI1 gene polymorphism modulating
18 glucocorticoid sensitivity was associated with a decline of the TLC at 3 years after LT,
19 and particularly in the recipients of CLT, the GLCCI1 variant was associated with a
20 decrease of the TLC and FEV1, and also a worse CLAD-free survival at 3 years after
21 CLT. Genotyping for this SNP might prove useful for adjusting the glucocorticoid dose to
22 prevent a decline of the TLC after LT and reduce the risk of CLAD after CLT.

23

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4

5 **Compliance with ethical standards**

6 **Conflict of Interest:** Haruchika Yamamoto and his co-authors have no conflicts of

7 interest.

8

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18

1 **Figure legends**

2 **Fig. 1. Postoperative changes of pulmonary function at 3 years after lung**
3 **transplantation (LT) in the recipients according to the GLCCI1 genotype.** (a) total
4 lung capacity (TLC), (b) forced expiratory volume in 1 second (FEV1), (c) forced vital
5 capacity (FVC) in all the recipients of LT. The percent baseline values of TLC in all the
6 recipients of LT (a) in the homozygous mutant T allele (TT) group were significantly
7 lower than those in the homozygous wild-type allele (CC) group.

8
9 **Fig. 2. Postoperative changes of pulmonary function at 3 years after cadaveric lung**
10 **transplantation (CLT) in the recipients according to the GLCCI1 genotype.** (a) total
11 lung capacity (TLC), (b) forced expiratory volume in 1 second (FEV1), (c) forced vital
12 capacity (FVC) in the recipients of CLT. The percent baseline values of TLC (a) and FEV1
13 (b) in the recipients of CLT in the homozygous mutant T allele (TT) group were
14 significantly lower than those in the homozygous wild-type allele (CC) group.

15
16 **Fig. 3. Chronic lung allograft dysfunction (CLAD)-free survival after lung**
17 **transplantation (LT) according to rs37972 genotype.** (a) There were no significant
18 differences in the CLAD-free survival among the three groups after LT. (b) In the
19 recipients of cadaveric lung transplantation (CLT), the CLAD-free survival in the
20 homozygous mutant T allele (TT) group was significantly worse than that in the
21 homozygous wild-type allele (CC) group (P = 0.016).

22