Acta Medica Okayama

Volume 53, Issue 5

1999 October 1999 Article 6

Modulatory effect of a serine protease inhibitor on surgical stress: its clinical implications.

Hiromi Iwagaki* Takahito Yagi[†] Naoto Urushihara[‡] Kenta Kobashi^{**} Yoshinori Morimoto^{††} Hiroshi Isozaki^{‡‡} Norihisa Takakura[§] Noriaki Tanaka[¶]

*Okayama University, †Okayama University, ‡Okayama University, **Okayama University, ††Okayama University, \$Okayama University, \$Okayama University,

Copyright ©1999 OKAYAMA UNIVERSITY MEDICAL SCHOOL. All rights reserved.

Modulatory effect of a serine protease inhibitor on surgical stress: its clinical implications.*

Hiromi Iwagaki, Takahito Yagi, Naoto Urushihara, Kenta Kobashi, Yoshinori Morimoto, Hiroshi Isozaki, Norihisa Takakura, and Noriaki Tanaka

Abstract

The relationship between endogenous cytokine antagonists and surgical stress is poorly understood. Surgical stress induces immunosuppression, and the reversed therapy of postoperative immunosuppression has been expected. The aim of the present study was to assess the effect of a serine protease inhibitor on postoperative immune reactivity. Twenty patients with colorectal cancer were randomly separated into experimental and control groups of 10 patients each. The experimental group received perioperative administration of a serine protease inhibitor while the control group did not. Plasma levels of cytokine antagonists, which suppress cell-mediated immunity, such as cortisol, interleukin-1 receptor antagonist, soluble interleukin-2 receptor (sIL-2R) and soluble tumor necrosis factors p55, p75 (sTNF-R55, -R75) were simultaneously measured. Significant reductions of plasma concentration of sIL-2R and sTNF-R55 were observed. Perioperative administration of a serine protease inhibitor may contribute to ameliorating immunosuppression after major surgery.

KEYWORDS: surgical stress, cytokine antagonist, protease inhibitor

*PMID: 10561733 [PubMed - indexed for MEDLINE] Copyright (C) OKAYAMA UNIVERSITY MEDICAL SCHOOL

Modulatory Effect of a Serine Protease Inhibitor on Surgical Stress: Its Clinical Implications

Hiromi Iwagaki*, Takahito Yagi, Naoto Urushihara, Kenta Kobashi, Yoshinori Morimoto, Hiroshi Isozaki, Norihisa Takakura and Noriaki Tanaka

First Department of Surgery, Okayama University Medical School, Okayama 700-8558, Japan

The relationship between endogenous cytokine antagonists and surgical stress is poorly understood. Surgical stress induces immunosuppression, and the reversed therapy of postoperative immunosuppression has been expected. The aim of the present study was to assess the effect of a serine protease inhibitor on postoperative immune reactivity. Twenty patients with colorectal cancer were randomly separated into experimental and control groups of 10 patients each. The experimental group received perioperative administration of a serine protease inhibitor while the control group did not. Plasma levels of cytokine antagonists, which suppress cellmediated immunity, such as cortisol, interleukin-1 receptor antagonist, soluble interleukin-2 receptor (sIL-2R) and soluble tumor necrosis factors p55, p75 (sTNF-R55, -R75) were simultaneously measured. Significant reductions of plasma concentration of sIL-2R and sTNF-R55 were observed. Perioperative administration of a serine protease inhibitor may contribute to ameliorating immunosuppression after major surgery.

Key words: surgical stress, cytokine antagonist, protease inhibitor

P ostoperative immune response is believed to be mediated by the proinflammatory cytokines interleukin (IL)-1 and tumor necrosis factor (TNF), and modulated by the naturally occurring specific antagonists of these cytokines (1). Postoperative production of TNF and IL-1 following major surgery has been studied by other researchers (2–5). In contrast, production of cytokine antagonists following surgical stress has not been extensively studied. Increased concentrations of soluble TNF receptor (sTNF-R), IL-1 receptor antagonist (IL-1Ra) and soluble IL-2 receptor (sIL-2R) have been reported in various pathological states, and are usually correlated with disease activity (6–8). Furthermore, recent reports have concluded that increased plasma concentrations of sTNF-R after major surgery are predictive of subsequent sepsis (9). The purpose of the present study was to investigate the serial changes of cytokine antagonists after surgery and the effect of perioperative administration of a serine protease inhibitor on postoperative production of cytokine antagonists.

Patients and Methods

Twenty patients were randomly separated into experimental and control groups of 10 patients each. The experimental group received a perioperative 24-h administration of intravenous serine protease inhibitor (FUT-175: 6-amidino-2-naphtyl p-guanidino-benzoate dimethanesulphonate; mol.wt. = 539.59, Torii Co. Ltd., Tokyo, Japan; protease inhibitor group). The control group received no protease inhibitor (control group). A serine protease inhibitor (FUT-175) was dissolved in 5 % glucose solution and injected intravenously. Patients in the experimental group were started on an intravenous drip of a protease inhibitor at a dose of 0.1 mg/kg/h immediately before surgery for resection of colorectal cancer. Administration of a protease inhibitor was continued at the same rate during surgery and for 24 h after the completion of surgery. No patient was suffering from any another malignancy, autoimmune, neurological or connective tissue disease. Nor were any of them taking steroid or antihypertensive medication. All patients gave informed

^{*} To whom correspondence should be addressed.

240 IWAGAKI ET AL.

written consent. Table 1 outlines the patient characteristics. There were no significant intergroup differences.

Venous blood samples for serological assays were taken at 8:00 a.m. each morning. Samples were obtained on the day of surgery (day 0) and on postoperative days 1, 2 and 7. Plasma levels of IL-1Ra, sIL-2R and sTNF-R55, -R75 were quantified by commercial immunosorbent assay (ELISA) kits (IL-1Ra and sTNF-R55, -R75, Amersham Life Science; sIL-2R, T Cell Diagnostics, Inc.), and serum cortisol levels were simultaneously measured by radioimmunoassay using Gamma-Coat-Cortisol radioimmunoassay kits (Travenol Co. Ltd.).

Data are reported as the relative mean. Between groups, data were compared at defined time points using the Wilcoxon 2-sample test. A P value less than 0.05 was regarded as significant.

Results and Discussion

Previous studies have demonstrated a marked increase in plasma IL-6 post-operatively, but have been unable to detect a consistent rise in plasma IL-1 β and TNF- α (10). Failure to detect IL-1 β or TNF- α in plasma may be due to the transient, local paracrine release of these agents and their rapid proteolytic degradation (11). Alternatively, the assays may not be sufficiently sensitive, or may fail to detect receptor-bound or protein-bound cytokines (12). In severe disease and trauma, cytokines produced in excess or for prolonged periods may cause local and systemic toxicity (13). Their biological activity is normally controlled by regulation of biosynthesis and release, with the effects limited by possible counteractive mechanisms: autoantibodies, inhibitory or opposing cytokines, and soluble receptors or receptor antagonists.

In the present study, significant elevations in plasma levels of cytokine antagonists such as cortisol, sIL-2R, sTNF-R55, -75 and IL-1Ra on day 1 were observed after colorectal resection in the control group (Fig. 1). These cytokine antagonists would be expected to effectively neutralize their respective ligands in serum, based on their behavior in vitro (14-16). However, the capacity of cytokine antagonists to effectively neutralize their respective ligands in humans has not been well established (17, 18). As mentioned ealier, a recent report has concluded that increased plasma concentrations of sTNF-R55 after major surgery are predictive of subsequent sepsis (9). These results suggest that levels of postoperative cytokine antagonist production may be excessive and over the amount necessary to achieve full ligand antagonism under in vivo conditions. It has been reported that surgical stress induces immunosuppression, the mechanism of which might be, at least in part, due to the induction of these cytokine antagonists.

IL-2 is produced by mature T-lymphocytes in response to lectin or antigen activation (19–21) and promotes the in vitro growth of T-lymphocytes by interaction with IL-2-specific cell surface receptors (IL-2R) that appear in a time-dependent manner on activated T-cells (22). sIL-2R is released from activated human lymphoid cells *in vitro*, and markedly elevated levels of plasma sIL-2R have recently been reported in patients with hematologic malignancy (23–25) and autoimmunne disorders such as systemic lupus erythematosus (26). The plasma level of sIL-2R represents a good marker of disease activity and, apparently, the activity of a T-cell subpopulation (25, 26). sIL-2R binds with IL-2, thus making less IL-2 available for binding with IL-2 cell surface receptors (27).

sTNF-R is released by activated neutrophils (28), mononuclear blood cells and fibroblasts (29), by shedding from cell membranes. A recent report found that increased plasma sTNF-R55 concentrations one day after major surgery were predictive of subsequent sepsis and poor outcome, and that sTNF-R75 was less predictive (9,

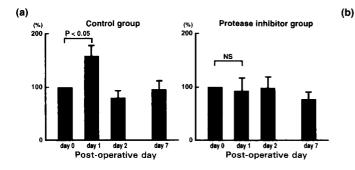
Table I Profile	of	patients
-----------------	----	----------

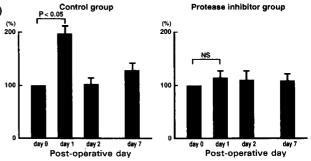
	Protease inhibitor group	Control group	Significance
Sex (n; male/female)	2/3	6/9	NS
Age (years)	61.2 ± 4.9	54.6 \pm 11.2	NS
Length of surgery (min)	257 ± 123.0	331.0 ± 136.3	NS
Blood loss (ml)	1319.0 \pm 2078.5	1453.3 ± 1462.5	NS
Dukes (B/C/D)	1/2/2	4/5/6	NS

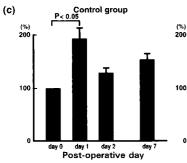
Values represent mean \pm SD. Statistical significance was analyzed as appropriate. NS: Not significant.

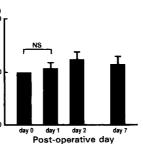
October 1999

Protease Inhibitor on Surgical Stress 241

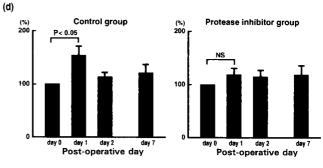








Protease inhibitor group



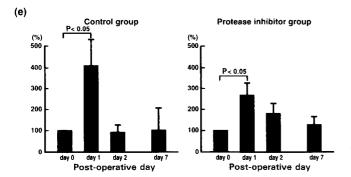


Fig. 1 The serial plasma levels of cortisol (a), soluble interleukin-2 receptor (b), soluble tumor necrosis factor receptor-type I (sTNF-R55) (c), soluble tumor necrosis factor receptor-type II (sTNF-R75) (d) and interleukin-1 receptor antagonist (e) following surgery with or without perioperative protease inhibitor treatment. NS: significant.

242 Iwagaki et al.

30). Furthermore, plasma sTNF-R55 concentrations measured after liver transplantation reportedly increase before the development of severe infection (31). Plasma sTNF-R55 concentrations were significantly higher in the infected group than in the uninfected group after liver resection (32). Bacterial products such as formyl methionyl leucyl phenylalanine may be the primary inducers of sTNF-R release from neutrophils in patients with bacterial infection (28). Major surgery is reported to impair gut barrier function, resulting in increased bacterial translocation (32). Therefore, intraoperative bacterial translocation from the bowel may be a major stimulus causing the increased plasma concentration of sTNF-R.

Human monocytes produce IL-1 β and IL-1Ra proteins in similar quantities (33). As monocytes mature to become macrophages, IL-1 β production is downregulated and IL-1Ra production is greatly enhanced (34). Thus, tissue macrophages may be the major source of IL-1Ra after surgery. As described above, immune suppression has long been considered a consequence of major surgery (35, 36). Considering the key role played by IL-1 in influencing the growth and differentiation of immunocompetent lymphocytes, it seems reasonable to assume that the immunological suppression often observed after major surgery might be related to the IL-1Ra response which we have observed in the present study. A recent report of the ability of IL-1Ra to down-regulate IL-2, IL-2R expression and lectin-stimulated lymphocyte proliferation (37), would lend further support to the idea that IL-1Ra plays a role in modulating immune response.

IL-6 is a pleiotropic cytokine involved in the regulation of immune responses and the acute-phase reaction (38). This cytokine is produced by a variety of cells after stimulation, such as infection, trauma or immunological challenge. Previous reports have also identified IL-6 as the earliest detectable cytokine response associated with major surgery (32, 39). Earlier studies showed that IL-6 inhibits LPS-induced TNF- α and IL-1 β production in cultured human monocytes, and in mice in vivo (40, 41), which suggest that IL-6 possesses antiinflammatory properties. A recent report also suggests that the antiinflammatory properties of IL-6 may be due to the induction of IL-1Ra synthesis and the release of sTNF-R (42). The same paper also suggests that tissue macrophages may be an important source of IL-6-induced cytokine antagonists.

In patients treated with perioperative administration of a protease inhibitor, with the exception of IL-1Ra, no significant increases of cytokine antagonists were observed (Fig. 1). The levels of IL-1Ra on day 1 in the protease inhibitor group were lower than those in the control group, although this was not statistically significant. The values of sIL-2R and sTNF-R55 in the protease inhibitor group on day 1 were significantly lower than those in the control group (Table 2). The low induction of soluble cytokine antagonists in the protease inhibitor group can be accounted for by two contrasting theories: The first holds that cytokine receptors are released from the cell surface through proteolytic cleavage, and therefore, that protease inhibitors may inhibit the shedding of cytokine receptors (43, 44). The second, described above, holds that IL-6 somehow causes the induction of cytokine antagonists such as IL-1Ra and sTNF-R (42). A serine protease inhibitor has been reported to cause a reduction in plasma concentration of IL-6,

 Table 2
 Comparison of plasma levels of cortisol, soluble interleukin-2 receptor (slL-2R), soluble tumor necrosis factor receptor type I (sTNF-R55), soluble tumor necrosis factor receptor type II (sTNF-R75) and interleukin-1 receptor antagonis(IL-1Ra) between the experimental group (protease inhibitor group) and the control group

		Protease inhibitor group	Control group	Significance
Cortisol	day O	100.0	100.0	
	1	91.6 ± 24.6	159.9 ± 20.8	NS
	2	98.9 ± 21.5	79.5 ± 18.0	NS
	7	$\textbf{78.8} \pm \textbf{16.2}$	$93.4\pm$ 19.3	NS
sIL-2R	day O	100.0	100.0	_
	1	$ 3.2 \pm 2. $	179.8 ± 16.5	P < 0.01
	2	107.5 ± 18.6	105.5 ± 6.9	NS
	7	108.2 ± 15.2	126.0 \pm 11.8	NS
sTNF-R55	day O	100.0	00.0	_
	1	104.2 ± 10.9	194.5 ± 22.3	P < 0.05
	2	120.6 \pm 16.5	125.9 ± 7.3	NS
	7	118.4 \pm 14.1	153.9 ± 9.8	NS
sTNF-R75	day O	100.0	100.0	
	1	118.5 ± 11.8	152.8 ± 18.2	NS
	2	116.9 \pm 13.6	114.4 ± 6.8	NS
	7	118.8 ± 18.6	120.7 ± 17.0	NS
IL-IRa	day 0	100.0	100.0	
	1	257.3 ± 64.0	406.8 ± 120.7	NS
	2	178.0 ± 44.8	$93.4\pm~37.0$	NS
	7	126.1 \pm 38.6	102.9 ± 101.4	NS

Values represent mean \pm SE. Wilcoxon 2-sample test was used to evaluate the statistical significance of intergroup differences.

October 1999

which, in turn, may reduce the plasma levels of cytokine antagonists.

It has been reported that patients who suffer postoperative complications have higher plasma levels of cytokine antagonists than those who do not suffer complications (30) and that there is a marked increase in plasma cytokine antagonists in patients at risk for developing multiple organ failure (45). The present research demonstrates that perioperative administration of a serine protease inhibitor yields a reduction in the postoperative induction of cytokine antagonists related to immunological depression. These results suggest that a protease inhibitor has protective effects against surgical stress induced immunosuppression.

References

- Davies MG and Hagen PO: Systemic inflammatory response syndrome. Br J Surg (1997) 84, 920–935.
- Baigrie RJ, Lamont PM, Kwiatkowski D, Dallman MJ and Morris PJ: Systemic cytokine response after major surgery. Br J Surg (1992) 79, 757-760.
- Foulds S, Cheshire NJ, Schachter M, Wolfe JH and Mansfield AO: Endotoxin related early neutrophil activation is associated with outcome after thoracoabdomnal aortic aneurysm repair. Br J Surg (1997) 84, 172-177.
- Wortel CH, van Deventer SJH, Aarden LA, Lygidakis NJ, Buller HR, Hoek FJ, Horikx J and ten Cate JW: Interleukin-6 mediates host defense responses induced by abdominal surgery. Surgery (1993) 114, 564–570.
- Kimura F, Miyazaki M, Suwa T, Kakizaki S, Itoh H, Kaiho T and Nakajina N: Increased serum interleukin-6 level and reduction of hepatic acute-phase response after major hepatectomy. Eur Surg Res (1996) 28, 96–103.
- 6. Van Zee KJ, Kohno T, Fischer E, Rock CS, Moldawer LL and Lowry SF: Tumor necrosis factor soluble receptors circulate during experimental and clinical inflammation and can protect against excessive tumor necrosis factor α *in vitro* and *in vivo*. Proc Natl Acad Sci USA (1992) **89**, 4845-4849.
- Aderka D, Wysenbeek A, Engelmann H, Cope AP, Brennan F, Molad Y, Hornik V, Levo Y, Maini RN and Feldmann M: Correlation between serum levels of soluble tumor necrosis factor receptor and disease activity in systemic lupus erythematosus. Arthritis Rheum (1993) 36, 1111-1120.
- Tilg H, Vogel W, Wiedermann CJ, Shapiro L, Herold M, Judmaier G and Dinarello CA: Circulating interleukin-1 and tumor necrosis factor antagonists in liver disease. Hepatology (1993) 18, 1132-1138.
- Pilz G, Fraunberger P, Appel R, Kreuzer E, Werdan K, Walli A and Seidel D: Early prediction of outcome in score-identified, postcardiac surgical patients at high risk for sepsis, using soluble tumor necrosis factor receptor-p55 concentrations. Crit Care Med (1996) 24, 596– 600.
- Cruickshank AM, Fraser WD, Burns HJ, Van Damme J and Shenkin A: Response of serum interleukin-6 in patients undergoing elective surgery of varying severity. Clin Sci (1990) 79, 161–165.
- 11. Tracey KJ, Morgello S, Koplin B, Fahey TJ III, Fox J, Aledo A,

Protease Inhibitor on Surgical Stress 243

Manogue KR and Cerami: Metabolic effects of cachectin/tumor necrosis factor are modified by site of production: Cachectin/tumor necrosis factor-secreting tumor in skeletal muscle induces chronic cachexia, while implantation in brain induces predominantly acute anorexia. J Clin Invest (1990) **86**, 2014–2024.

- Engelberts I, Stephens S, Francot GJ, van der Linden CJ, Buurman WA: Evidence for different effects of soluble TNF-receptors on various TNF measurements in human biological fluids. Lancet (1991) 338, 515–516.
- Tracey KJ, Wei H, Manogue KR, Fong Y, Hesse DG, Nguyen HT, Kuo GC, Beutler B, Cotran RS, Cerami A and Lowry SF: Cachectin/tumor necrosis factor induces cachexia, anaemia, and inflammation. J Exp Med (1988) 167, 1211–1227.
- Hale KK, Smith CG, Baker SL, Vanderslice RW, Squires CH, Gleason TM, Tucker KK, Kohno T and Russell DA: Multifunctional regulation of the biological effects of TNF-alpha by the soluble type I and type II TNF receptors. Cytokine (1995) 7, 26–38.
- Granowitz EV, Clark BD, Vannier E, Callahan MV and Dinarello CA: Effect of interleukin-1 (IL-1) blockade oncytokine synthesis: I. IL-1 receptor antagonist inhibits IL-1-induced cytokine synthesis and blocks the binding of IL-1 to its type II receptor on human monocytes. Blood (1992) 79, 2356–2363.
- Granowitz EV, Vannier E, Poutsiaka DD and Dinarello CA: Effect of interleukin-I (IL-I) blockade on cytokine synthesis: II. IL-I receptor antagonist inhibits lipopolysaccharide-induced cytokine synthesis by human monocytes. Blood (1992) 79, 2364–2369.
- Fisher CJ Jr, Agosti JM, Opal SM, Lowry SF, Balk RA, Sadoff JC, Abraham E, Sohein RM and Benjamin E: Treatment of septic shock with the tumor necrosis factor receptor: Fc fusion protein. Ths Soluble TNF Receptor Sepsis Study Group. N Engl J Med (1996) 334, 1697-1702.
- Opal SM, Fisher CJ Jr, Dhainaut JF, Vincent JL, Brase R, Lowry SF, Sadoff JC, Slotman GJ, Levy H, Balk RA, Shelly MP, Pribble JP, La Brecque JF, Lookabaugh J, Donovan H, Dubin H, Baughman R, Norman J, De Maria E, Matzel K, Abraham E and Seneff M: Confirmatory interleukin-1 receptor antagonist trial in severe sepsis: A phase III, randomized, double-blind, placebo-controlled, multicenter trial. The Interleukin-1 Receptor Antagonist Sepsis Investigator Group. Crit Care Med (1997) 25, 1115–1123.
- Robb RJ, Munck A and Smith KA: T cell growth factor receptors: Quantitation, specificity, and biologic relevance. J Exp Med (1981) 154, 1455–1474.
- Robb RJ: Interleukin-2: The molecule and its function. Immunol Today (1984) 5, 203–209.
- 21. Smith KA: Interleukin-2. Annu Rev Immunol (1984) 2, 319-333.
- Cantrell DA and Smith KA: Transient expression of interleukin-2 receptors: Consequences for T cell growth. J Exp Med (1983) 158, 1895-1911.
- Waldmann TA: The structure, function and expression of interleukin-2 receptor on normal and malignant lymphocytes. Science (1986) 32, 727–732.
- Jung LKL, Hana T and Fu SM: Detection and functional studies of TAC antigen on activated human B cells. J Exp Med (1984) 160, 1597– 1605.
- Harrington DS, Patil K and Lai PK: Soluble interleukin-2 receptors in patients with malignant lymphoma. Arch Pathol Lab Med (1988) 112, 597-601.
- Semenzato G, Bambara L and Biasi D: Increased serum levels of soluble interleukin-2 receptor in patients with systemic lupus erythematosus and rheumatoid arthritis. J Clin Immunol (1988) 8, 447-451.
- 27. Rubin LA, Jay G and Nelson DL: The released interleukin-2 receptor

244 IWAGAKI ET AL.

binds interleukin-2 efficiently. J Immunol (1986) 137, 3841-3845.

- Porteu F and Nathan C: Shedding of tumor necrosis factor receptors by activated human neutrophils. J Exp Med (1990) 172, 599-607.
- Lantz M, Gullberg U, Nilsson E and Olsson I: Characterization in vitro of a human tumor necrosis factor-binding protein: a soluble form of a tumor necrosis factor receptor. J Clin Invest (1990) 86, 1396–1402.
- Kimura F, Miyazaki M, Suwa T, Sugiura T, Shinoda T, Itoh H, Ambiru S, Shimizu H and Nakagawa K: Plasma concentration of cytokine antagonists in patients with infection following liver resection. Br J Surg (1998) 85, 1631–1635.
- Mueller AR, Platz KP, Wiehe I, Monticelli F, Lierath J and Keitel M: Cytokine pattern in patients with infections after liver transplantation. Transpl Int (1996) 9, 126-131.
- Wang XD, Parsson H, Andersson R, Soltesz V, Johansson K and Bengmark S: Bacterial translocation, intestinal ultrastructure and cell membrane permeability early after major liver resection in the rat. Br J Surg (1994) 81, 579–584.
- Arend WP, Smith MF Jr, Janson RW, Joslin FG: IL-I receptor antagonist and IL-Iβ production in human monocytes are regulated differently. J Immunol (1991) 147, 1530–1536.
- Janson RW, Joslin FG, Arend WP: The effects of differentiating agents on IL-1β production in cultured human monocytes. J Immunol (1990) 145, 2161–2166.
- Salo M: Effects of anaesthesia and surgery on the immune response; in Trauma, Stress and Immunity in Anaesthesia and Surgery, Watkins J and Salo M eds, Butterworths, London (1982) pp 211-253.
- Green DR and Faist E: Trauma and the immune response. Immunol Today (1988) 9, 253-255.

ACTA MED OKAYAMA VOI. 53 No. 5

- Conti P, Panara MR and Porrini AM: Inhibition of interleukin-1 (alpha and beta), interleukin-2 secretion and surface expression of interleukin-2 receptor (IL-2R) by a novel cytokine interleukin-1 receptor antagonist (IL-1Ra). Scand J Immunol (1992) 36, 27–33.
- Kishimoto T, Akira S and Taga T: Interleukin-6 and its receptor: A paradigm for cytokines. Science (1992) 258, 593.
- Sweed Y, Puri P and Reen DJ: Early induction of IL-6 in infants undergoing major abdominal surgery. J Pediat Surg (1992) 27, 1033 -1037.
- Schindler R, Mancilla J, Endres S, Ghorbani R, Clark SC and Dinarello CA: Correlations and interactions in the production of interleukin-6 (IL-6), IL-1 and tumor necrosis factor (TNF) in human blood mononuclear cells: IL-6 suppresses IL-1 and TNF. Blood (1990) 75, 40-47.
- Aderka D, Le JM and Vilcek J: IL-6 inhibits lipopolysaccharide-induced tumor necrosis factor production in cultured human monocytes, U937 cells, and in mice, J Immunol (1989) 143, 3517–3523.
- Tilg H, Trehu E, Atkins MB, Dinarello CA and Mier JW: Interleukin-6 (IL-6) as an anti-ilammatory cytokine: Induction of circulating IL-1 receptor antagonist and soluble tumor necrosis factor receptor p55. Blood (1994) 183, 113-118.
- Robb RJ and Kutny RM: Structure-function relationships for the IL2receptor system: IV. Analysis of the sequence and ligand-binding properties of soluble Tac protein. J Immunol (1987) 139, 855–862.
- Iwagaki H, Hizuta A, Iwadou H, Perdomo JA, Tanaka N and Orita K: Clinical value of soluble interleukin-2 receptor in infectious complications. Acta Med Okayama (1994) 48, 225-226.
- Iwagaki H, Hizuta A, Uomoto M, Takeuchi Y, Kohka H, Okamoto T and Tanaka N: Clinical value of cytokine antagonists in infectious complications. Res Commun Mol Pathol Pharmacol (1997) 96, 25–34.

Received February 23, 1999; accepted June 29, 1999.